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Modelling Insect Natality Using Splines

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SUMMARY

We present methodology for analyzing age-specific birth rates, with particular emphasis on describing variability within a cohort of like-aged individuals. We use a model structure appropriate for natality in insects and other arthropod species. Our approach separates natality into a baseline rate representative of the effects of the environment for the population and a factor that represents the unique effect of each individual in the cohort. We employ smoothing splines with the penalized likelihood methodology for estimation. We illustrate the methodology by applying it to data on leafhopper egg-laying rates.

1. Introduction

The age-specific natality schedule provides information of fundamental importance in animal population ecology. Conventional methods for studying population dynamics [e.g., the Leslie matrix model (Plant, 1986)] require information on age-specific natality for the population, but for the most part these approaches require only population averages and ignore any associated variability. However, there has been a growing interest in exploring the implications that variability in vital rates (birth and death) has for overall population behavior (Meyer et al., 1986; Slade and Levenson, 1982; Strong, 1986). Arthropods in general, and certain insect species in particular, exhibit considerable variation in age-specific birth rates and thus make good candidates for investigating individual variability and its effects on population dynamics.

A common approach in insect natality studies considers the cumulative number of births as a function of time for an individual as

$$N(t) = \int_0^t m(x) \ dx$$

with m(t) the observed birth rate. The expected birth rate, h(t) = Em(t), is then modelled with some parametric form, such as a logistic, exponential, or Gompertz curve (cf. Madden, 1985). In contrast, we develop a nonparametric method for modelling insect natality that allows us to characterize an overall or "baseline" birth rate and to analyze individual contributions to that rate. Our starting point considers the number of eggs laid per unit time as a nonstationary Poisson process (Cinlar, 1975), and we propose to estimate the

age-specific Poisson rates by penalized maximum likelihood using spline-based generalized linear models.

The model structure we propose embodies the general characteristics of insect natality. In addition to high individual variability, these include strong dependency on age and sensitivity to environmental conditions (especially temperature). Let $h^0(t)$ be the birth rate mediated by the common environment, the age-specific "baseline rate" common to all individuals subject to the same conditions. Furthermore, we suppose that variability introduced by individual differences over time (= age) affects the birth rate in a multiplicative fashion. Let $a_i(t)$ be the "individual factor" that depends on unique characteristics during the life of individual i. We consider modelling the birth rate for the ith individual, $h_i(t)$, as

$$h_i(t) = h^0(t)a_i(t). (1)$$

That is, individual-to-individual variation has a relative effect on the baseline rate. Other models, such as an additive one where individual factors have an absolute effect on the number of births, are possible but not considered here. The model structure (1) has been considered extensively for mortality rates (Cox, 1972). If individual birth rate curves $h_i(t)$ follow the same shape as the baseline birth rate (on a log scale), then $a_i(t)$ is constant. If not, changes in $a_i(t)$ reflect time-dependent changes in individual natality (e.g., due to physiological changes). Taking logs in (1) and defining $\theta_i = \log h$, $\theta^0 = \log h^0$, and $\alpha_i = \log \alpha_i$, we have

$$\theta_i(t) = \theta^0(t) + \alpha_i(t). \tag{2}$$

Bartoszyński et al. (1981) and Clevenson and Zidek (1977) considered such a model for Poisson data with $\alpha_i(t)$ assumed constant. This model is a special case of the generalized additive model (Hastie and Tibshirani, 1986).

We estimate the log baseline rate $\theta^0(t)$ and individual factors $\alpha_i(t)$ as smoothly changing functions of time. Our method requires the imposition of some measure of desired smoothness on the estimates through a "penalty" on overall curvature of the functions. We maximize the likelihood plus the penalty, striking a compromise between fidelity to the data (with rather bumpy estimates) and simplicity in curve shape (with straight-line estimates). This maximum penalized likelihood methodology has received growing attention recently for graphical examination of smoothly changing curves (cf. O'Sullivan, Yandell, and Raynor, 1986; Green and Yandell, 1985; Silverman, 1985; and Hastie and Tibshirani, 1986).

We outline the methodology in Section 2. We demonstrate the utility of our approach by applying it to leafhopper data sets: baseline rates are analyzed in Section 3, and individual variability is examined in Section 4. We make some conclusions and remarks in Section 5.

2. Splines and Penalized Likelihoods

We present here our estimation procedure for the baseline birth rate and the individual factors using the penalized likelihood method. Added detail is presented in the Appendix. We consider first a single female and for notational simplicity drop the individual subscript i. Thus, our goal is estimation of h(t) or, equivalently, $\theta(t) = \log h(t)$. Suppose the female is observed at n distinct times, $0 < t_1 < \cdots < t_n$, with $t_0 = 0$ being the time of adult emergence. During an interval between observation times, say t_{j-1} and t_j , one would expect to see H_j births, with

$$H_{j} = \int_{t_{j-1}}^{t_{j}} h(t) dt.$$
 (3)

The number of births occurring during this interval, m_j , should have approximately a Poisson distribution with mean H_j . The best (maximum likelihood) estimate of H_j is m_j , the observed count, and the best estimate of h(t) for t between t_{j-1} and t_j is

$$\hat{h}(t) = m_i/d_i, \quad \text{with} \quad d_i = t_{i-1} - t_i.$$
 (4)

Unfortunately, if there are many short intervals between observations, this (nonsmoothed) estimate of birth rate $\hat{h}(t)$ ends up being very rough.

The basic idea in our approach using smoothing splines is analogous to attaching a flexible rod to the crude estimates of $\theta(t) = \log h(t)$ from (4) at the "nodes" t_i ,

$$\hat{\theta}(t_i) = \log(m_i/d_i),\tag{5}$$

using springs that move vertically. Strong springs force the rod (or curve) to adhere to the point estimates, whereas weak springs lead to a linear estimate. Springs with intermediate strength allow the rod to form a smooth curve. Our approach leads to a curve that approximately follows a cubic polynomial between nodes. We use maximum likelihood imposing a "penalty" on rough curves, with a large penalty corresponding to a weak spring.

For a single individual, the Poisson distribution

$$\Pr\{m_j \mid H_j\} = (H_j)^{m_j} \exp(-H_j)/H_j!$$

leads to the log-likelihood based on observed counts m_1, \ldots, m_n , of

$$L = \log \prod_{j=1}^{n} \left(\frac{\Pr\{m_j \mid H_j\}}{\Pr\{m_j \mid m_j\}} \right) = \sum_{j=1}^{n} m_j (\log H_j - \log m_j) + \sum_{j=1}^{n} (m_j - H_j).$$
 (6)

The second sum can always be made 0 by setting $\Sigma H_j = \Sigma m_j$, and is ignored below. If one assumes that h is "smooth," one can allow the data to indicate what shape h has without specifying a parametric form. Approximating the integral H_j by $h(t_j)d_j$ (see Appendix) and using $\theta = \log h$, we can write the log-likelihood as

$$L = \sum_{j=1}^{n} m_{j} [\theta(t_{j}) + \log(d_{j}) - \log(m_{j})].$$
 (7)

Maximizing L directly leads to the estimates (5). Instead, we maximize the penalized likelihood,

$$L - \frac{1}{2}\lambda J(\theta), \tag{8}$$

with $J(\theta)$ the roughness penalty and λ an appropriate "spring constant." With a penalty $J(\theta)$ on overall curvature,

$$J(\theta) = \int [\theta''(t)]^2 dt, \tag{9}$$

the maximum penalized likelihood estimate is cubic between observation times but is continuous with a continuous derivative over the domain of observation. Other choices of the penalty J are possible, e.g., to emphasize a more Bayesian interpretation of $\theta(t)$ (Silverman, 1985; Wahba, 1985).

If the spring constant λ is near 0 (strong springs), the estimate reduces to the rough maximum likelihood estimate (4). A large value (weak springs) forces the estimate to be a straight line. An intermediate value of λ can be chosen by generalized cross validation (GCV) (Craven and Wahba, 1979), which compares each count (m_i) with the curve estimate obtained by excluding that count. Since nonparametric curve estimates are biased, one must strike a balance between bias and variance reduction. Selecting that λ which minimizes the GCV criterion tends to achieve this in the sense of approximately minimizing the expected mean squared error.

Suppose one observes r individuals, each with its own birth rate according to the model (1), and that m_{ij} offspring are found for female i at time t_j . The overall penalized likelihood can be written, using (5), as

$$L = \sum_{i=1}^{r} \sum_{j=1}^{n} m_{ij} [\theta^{0}(t_{j}) + \alpha_{i}(t_{j}) + \log d_{j} - \log m_{ij}] - P$$

where P incorporates the penalty. This can be expressed as

$$L = L(\theta^0) + \sum_{i=1}^r L(\alpha_i) - P,$$

with

$$L(\theta^{0}) = \sum_{j=1}^{n} m_{+j} [\theta^{0}(t_{j}) + \log(rd_{j}) - \log m_{+j}],$$

$$L(\alpha_{i}) = \sum_{j=1}^{n} m_{ij} [\alpha_{i}(t_{j}) + \log(m_{+j}/r) - \log m_{ij}], \quad i = 1, ..., r,$$
(10)

and $m_{+j} = \sum_i m_{ij}$. Note that $L(\theta^0)$ is a Poisson likelihood, and at time t_j , $\{m_{ij}\}_{i=1}^r$ is multinomial conditional on m_{+j} . Thus, one now has (r+1) log-likelihoods in (10).

Our earlier discussion of roughness focused on a single function, $\theta(t)$, for a single individual. We now have (r + 1) functions, $\theta^0(t)$ and $\alpha(t) = \{\alpha_i(t)\}_{i=1}^r$. We postulate a smooth model that allows for two distinct roughness penalties, one for the baseline rate $\theta^0(t)$ and the other jointly for the individual factors $\alpha(t)$. Thus, we need to specify two spring constants, λ_1 and λ_2 , one for each penalty. The resulting penalized likelihoods are

$$L(\theta^0, \lambda_1) = L(\theta^0) - \frac{1}{2}\lambda_1 J(\theta^0),$$
 (11)

$$L(\boldsymbol{\alpha}, \lambda_2) = \sum_{i=1}^r L(\alpha_i) - \frac{1}{2}\lambda_2 \sum_{i=1}^r J(\alpha_i).$$
 (12)

Recall that if λ_2 is large (weak springs), the individual factors α_i are forced to be nearly linear.

The penalized likelihood $L(\theta^0, \lambda_1)$ can be solved for fixed λ_1 using a Newton-Raphson iterative scheme with Fisher scoring typical of generalized linear models (McCullagh and Nelder, 1983; cf. Green, 1984). A similar argument holds for $L(\alpha, \lambda_2)$. At convergence, the deviances $D(\hat{\theta}^0) = -2L(\hat{\theta}^0)$ and $D(\hat{\alpha}) = -2\sum_{i=1}^r L(\hat{\alpha}_i)$ have approximately χ^2 distributions (Yandell and Green, 1986). One can estimate the degrees of freedom for each statistic and use these for testing. For instance, comparing $D(\hat{\alpha})$ with the deviance for constant α_i tests whether birth rates can be considered parallel (on a log scale), with a constant relative difference. Details are presented in the Appendix.

Algorithms for solving this problem can be found in Green and Yandell (1985) based on the B-spline algorithm of Hutchinson and de Hoog (1985). This is currently being prepared for general distribution in FORTRAN77. A multidimensional algorithm based on GCVPACK (Bates et al., 1987) appears in Yandell (1988) and is available in FORTRAN77 from the author.

3. Potato Leafhoppers and Temperature

Hogg (1985) studied population dynamics of the potato leafhopper, an important crop pest, in controlled environments at the University of Wisconsin–Madison Biotron. Three temperature regimes were established with daily fluctuations designed to follow the cycle typically found under "cool," "intermediate," and "warm" summer field conditions in

southern Wisconsin. Analytical methods included age-specific life tables and Leslie matrix modelling, but did not incorporate individual variation in birth rates.

Here, we estimated the baseline oviposition rate h^0 for leafhoppers at each of the three temperature regimes. Each regime had 25 or 26 females observed three times a week from adult emergence to death or removal from study. Of these, some never produced eggs and were not further considered, leaving 24, 21, and 23 individuals for cool, intermediate, and warm regimes, respectively. We based estimates on data from the age of first egg-laying through the last interval in which eggs were laid. As a consequence, the population of each group could change over the course of the study as individuals died or were lost from the study.

We estimated the baseline rates using (11). The smoothing parameters were chosen using GCV and gave fairly smooth estimates (Figure 1). Note that the cool regime baseline rate

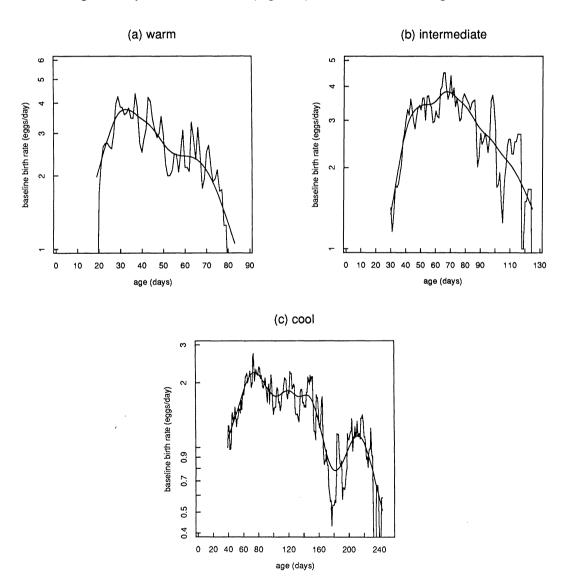


Figure 1. Curve estimates for potato leafhopper baseline oviposition rates for (a) warm, (b) intermediate, and (c) cool regimes. Smooth lines are penalized estimates; rough lines are interpolated daily estimates.

estimate has more structure to it, reflecting the fact that there were two or three times as many records for this regime because leafhoppers tended to live longer at lower temperatures. The baseline rate curves all tend to exhibit an early peak in oviposition rate, followed by a gradual decline.

Potato leafhopper longevity is greatly influenced by the environment (note the different time scales in Figure 1). One might ask whether the intermediate and warm regime baseline rates are accelerated versions of the cool regime baseline rate. To examine this, we converted time to "physiological time" units of degree-days, which combine calendar time and temperature to reflect more realistically the leafhoppers' growth and maturation (Figure 2). It appears that the shapes of the baseline rate curves are roughly similar, supporting the notion of an accelerated time scale. However, there are differences in magnitude of oviposition rates among the three regimes. In particular, on a physiological time scale, the intermediate regime appears to be the most favorable for oviposition (cf. Hogg, 1985).

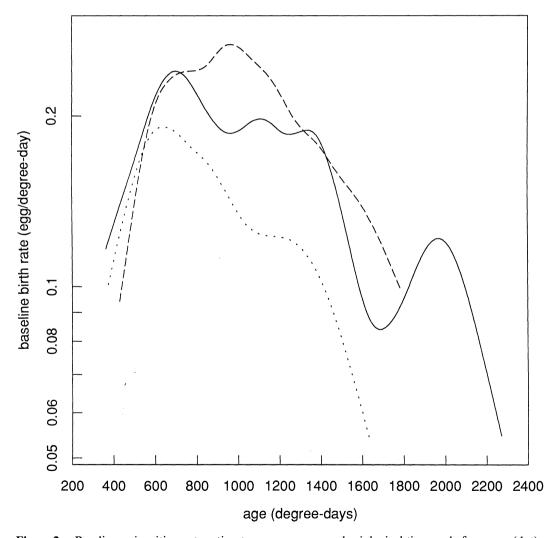


Figure 2. Baseline oviposition rate estimates on a common physiological time scale for warm (dot), intermediate (dash), and cool (solid) regimes.

4. Examining Individual Variability

We estimated individual factors a_i in each of the three temperature regimes using (12). We present here only results for the cool regime (Figure 3) since we have longer records for these individuals. Results for the other two regimes were similar.

Our main interest was in exhibiting the individual factors to look for interesting patterns not easily detected in the raw data or baseline rates. There were several individuals that had increased birth rates relative to the others early, but then died at a relatively early age. Note that some individuals had a much lower a_i in midlife (15, 20, 24), while some were just the opposite (18, 23).

We constructed deviance tests for constant individual factors to see whether the individual oviposition rates are parallel on a log scale, meaning that individual relative differences in egg-laying are constant over time. The tests were performed by comparing the difference in deviance between the constant a_i model and the smooth (spline) a_i model to a χ^2

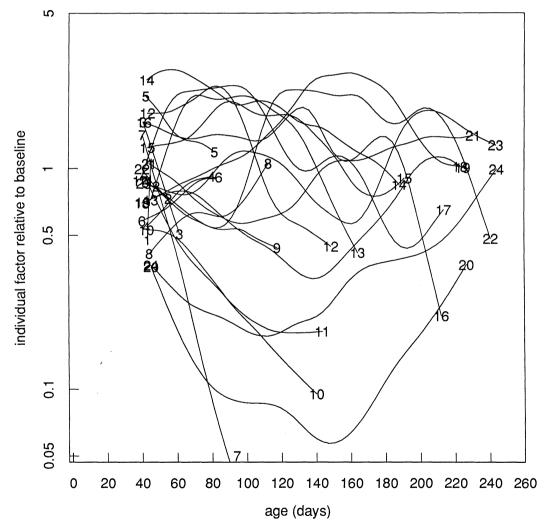


Figure 3. Variability among individual oviposition curves for 24 leafhoppers in the cold regime. Each curve is the individual factor, identified by survival rank.

distribution with degrees of freedom being the difference in degrees of freedom between the two models (Table 1). Note that the degrees of freedom are fractional and the test is approximate (Yandell and Green, 1986). These tests indicate that the individual factors are not constant; this result was consistent for all three regimes. Thus, the smoothly changing nature of the individual factors appears to be real, and marked differences exist in individual oviposition rates over time among leafhoppers in the same controlled environment.

Table 1
Deviance and degrees of freedom (in parentheses) for individual factors

	Model		
Regime	Constant	Spline	Difference
Warm	265.90	145.70	120.20
	(211)	(177.81)	(33.19)
Intermediate	856.55	614.31	242.24
	(436)	(393.31)	(42.69)
Cool	1,727.07	974.87	752.20
	(965)	(861.91)	(103.09)

Deviances are approximately χ^2 with d.f. which may be fractional. Difference is test of smooth spline against constant individual factor model.

5. Discussion

We have shown that it is possible to separate a baseline rate from individual factors that may modulate this baseline. We can examine pictures of the baseline rates and the individual factors, compare them visually, and even test for apparent patterns. While we have developed our model specifically for insect natality, the approach promises broader applicability for examining complicated counting data in other biological systems.

Different egg-laying patterns were exhibited by the potato leafhoppers within each regime, even among the longest-lived leafhoppers. However, we do not know if these represent distinct oviposition "strategies" within the leafhopper population, or whether they are simply a random sample from all possible patterns. These results suggest further work aimed at understanding the basis of individual variation in insect age-specific birth rates. It also suggests that population characteristics may not adequately reflect individual characteristics. For instance, the population net reproduction rate (cf. Pianka, 1978),

$$R^0 = \int_0^\infty l(t)h^0(t) \ dt,$$

with l(t) the survival probability, may differ markedly from the individual net reproduction rates

$$R_i = \int_0^\infty l(t)h_i(t) dt.$$

This net reproduction rate R^0 plays a key role in evolutionary theory (Williams, 1966).

Some aspects of the theory need refinement, notably demonstrating conclusively the distributions of tests and properties of estimates for semiparametric GLMs. However, as it stands, the methods presented here can be used for exploratory investigation to examine temporal and individual variability, and to suggest particular parametric models that may be useful for subsequent experiments. New questions about temporal patterns and

individual variability will emerge as this methodology is incorporated into statistical packages accessible to biologists.

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RÉSUMÉ

Nous présentons une méthodologie d'analyse dex taux de natalité âge-spécifiques, en insistant particulièrement sur la description de la variabilité au sein d'une cohorte d'animaux d'âges voisins. Nous utilisons une structure de modèle appropriée à la natalité des insectes et d'autres arthropodes. Notre approche sépare la natalité en un niveau de base représentatif des effets de l'environnement sur la population, et en un facteur représentant l'unique effet de chaque individu de la cohorte. Nous employons conjointement les fonctions de lissage splines et une méthode de vraisemblance pénalisée pour l'estimation. Nous illustrons cette méthodologie en l'appliquant à des données sur le taux d'oviposition des cicadelles.

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APPENDIX

We first justify our approximation of the integral (3) by the mean value theorem, and then present details of the maximum penalized likelihood estimation procedure for θ^0 , followed by a brief discussion of test statistics and the choice of spring constant λ_1 . Brief mention is made of estimating α .

The mean value theorem ensures that $H_j = h(t_j^*)(t_j - t_{j-1}) = h(t_j^*)d_j$ for some $t_{j-1} \le t_j^* \le t_j$. In practice, we assume that $t_j^* \approx (t_j + t_{j-1})/2$, but we ignore this below and treat t_j^* as t_j . Note that this approximation introduces some bias that could be important if there are only a small number of observation times. Using $\theta = \log h$ and the mean value theorem gives us (7).

Initial investigations showed that mortality in the population affects the baseline rates but has only a modest effect on the individual factors. One can adjust for mortality by modifying the population, and even allowing the baseline rate to have jumps (Shiau, Wahba, and Johnson, 1986). However, this has not been treated here.

The log-likelihood $L(\theta^0)$ can be approximated near its maximum, using a second-order Taylor series expansion, by a weighted least squares. Given an initial estimate θ_1^0 of θ^0 ,

$$L(\theta^{0}) \approx L(\theta_{1}^{0}) + \sum [\theta^{0}(t_{j}) - \theta_{1}^{0}(t_{j})](m_{+j} - w_{j}) - \frac{1}{2} \sum w_{j}[\theta^{0}(t_{j}) - \theta_{1}^{0}(t_{j})]^{2}$$

$$= L(\theta_{1}^{0}) + \frac{1}{2} \sum w_{i}(m_{+i} - w_{i})^{2} - \frac{1}{2} \sum w_{i}[v_{i} - \theta^{0}(t_{i})]^{2}, \qquad (13)$$

with "working-weights" and "working-values" being, respectively,

$$w_j = rd_j \exp[\theta_1^0(t_j)]$$
 and $y_j = \theta_1^0(t_j) + (m_{+j} - w_j)/w_j$. (14)

Note that given θ_1^0 only the last term of (13) depends on the unknown θ^0 .

The problem of finding θ^0 to maximize $L(\theta^0, \lambda_1)$ is equivalent to finding θ^0 to minimize

$$\sum w_{i} [y_{i} - \theta^{0}(t_{i})]^{2} + \lambda_{1} J(\theta^{0})$$
 (15)

and iterating to convergence. Typically one makes an initial guess at θ_1^0 , assigns w_j and y_j in (14), solves for a new θ^0 in (15), replaces θ_1^0 by the estimated θ^0 , and updates w_j and y_j , and so on, until the estimates converge adequately.

The estimate that minimizes (15) for penalty (9) can be written as (cf. Bates et al., 1987)

$$\theta^{0}(t) = \beta_{0} + \beta_{1}t + \beta_{2}t^{2} + \sum_{i=1}^{n} \delta_{i}|t - t_{i}|^{3}$$

with the condition that

$$\sum_{j=1}^{n} \delta_{j} t_{j}^{k} = 0, \quad k = 0, 1, 2.$$

To actually solve for this, we can write the penalty (9) in a quadratic form in the δ_i 's.

$$J(\theta^{0}) = \sum_{j=1}^{n} \sum_{k=1}^{n} \delta_{j} \delta_{k} |t_{j} - t_{k}|^{3},$$

allowing us to restate (15) as a quadratic form in the δ_i 's and β_k 's, which can be minimized by standard methods.

At convergence, the estimate can be written as

$$\hat{\boldsymbol{\theta}}^{0} = \mathbf{A}\mathbf{v},$$

with $\hat{\theta}^0$ and y vectors of the parameter estimates and working values, respectively, and A the matrix of coefficients. The "effective number of parameters" are trace (A) (Craven and Wahba, 1979). The deviance $D^2(\hat{\theta}^0) = -2L(\hat{\theta}^0)$ has approximately a χ^2 distribution with trace(I – A) degrees of freedom (often a fraction), which can be used for approximate tests (Yandell and Green, 1986).

The choice of λ_1 can be made by minimizing the GCV function,

$$V(\lambda_1) = n[\text{trace}(\mathbf{I} - \mathbf{A})]^{-2} \sum_{j=1}^{n} w_j [y_j - \hat{\theta}^0(t_j)]^2.$$
 (16)

Since w_j and y_j depend on the estimate $\hat{\theta}^0(t_j)$, one repeats the above minimization for several choices of λ and chooses that value which minimizes (16) by a grid search.

Similar arguments can be made for α and λ_2 . However, when data are not collected on the same days for all insects one must estimate the total number of offspring per each observation period on individual i. This can be done by linear interpolation, smoothing out the births for each individual, and replacing m_{+j} by the interpolated value in $L(\alpha_i)$ of (10). The penalty is a sum of terms such as those given above for $J(\theta^0)$. One then proceeds to the weighted least squares iteration to convergence. The deviance under the null model of constant a_i is simply $D^2(\bar{\alpha}) = -2 \sum_{i=1}^r L(\bar{\alpha}_i)$ from (10) with $\alpha_i(t_j)$ replaced by $\bar{\alpha}_i = \log(m_{i+}/m_{++})$, the maximum likelihood estimator for constant individual factors. The difference in deviances reported in Table 2 is simply $D^2(\bar{\alpha}) - D^2(\hat{\alpha})$.