

QTLs & Microarrays Overview

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- what is a QTL?
- what is the goal of your QTL study?
- why worry about multiple QTL?
- why study multiple traits together?

what is a QTL?

- QTL = quantitative trait locus (or loci)
 - trait = phenotype = characteristic of interest
 - quantitative = measured somehow
 - qualitative traits can often be directly mapped
 - quantitative traits not readily mapped
- locus = location in genome affecting trait
 - gene or collection of tightly linked genes
 - some physical feature of genome

what is the goal of QTL study?

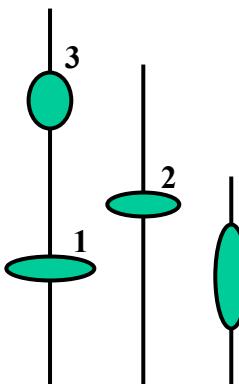
- uncover underlying biochemistry
 - identify how networks function, break down
 - find useful candidates for (medical) intervention
 - epistasis may play key role
 - statistical goal: maximize number of correctly identified QTL
- basic science/evolution
 - how is the genome organized?
 - identify units of natural selection
 - additive effects may be most important (Wright/Fisher debate)
 - statistical goal: maximize number of correctly identified QTL
- select “elite” individuals
 - predict phenotype (breeding value) using suite of characteristics (phenotypes) translated into a few QTL
 - statistical goal: minimize prediction error

why worry about multiple QTL?

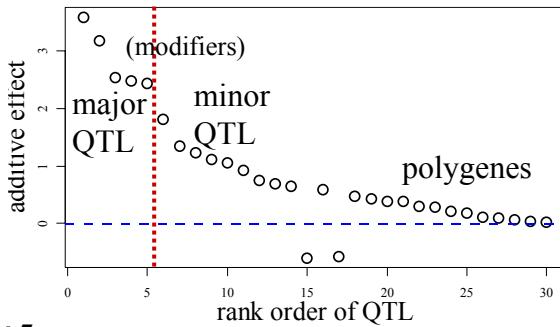
- so many possible genetic architectures!
 - number and positions of loci
 - gene action: additive, dominance, epistasis
 - how to efficiently search the model space?
- how to select “best” or “better” model(s)?
 - what criteria to use? where to draw the line?
 - shades of gray: exploratory vs. confirmatory study
 - how to balance false positives, false negatives?
- what are the key “features” of model?
 - means, variances & covariances, confidence regions
 - marginal or conditional distributions

Pareto diagram of QTL effects

major QTL on
linkage map



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advantages of multiple QTL approach

- improve statistical power, precision
 - increase number of QTL detected
 - better estimates of loci: less bias, smaller intervals
- improve inference of complex genetic architecture
 - patterns and individual elements of epistasis
 - appropriate estimates of means, variances, covariances
 - asymptotically unbiased, efficient
 - assess relative contributions of different QTL
- improve estimates of genotypic values
 - less bias (more accurate) and smaller variance (more precise)
 - mean squared error = $MSE = (\text{bias})^2 + \text{variance}$

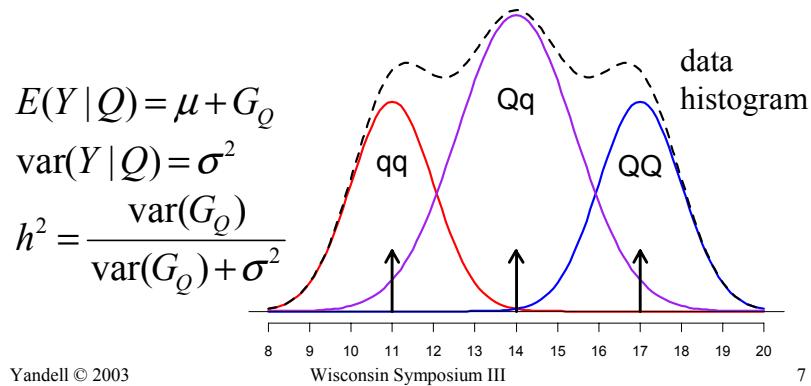
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typical phenotype assumptions

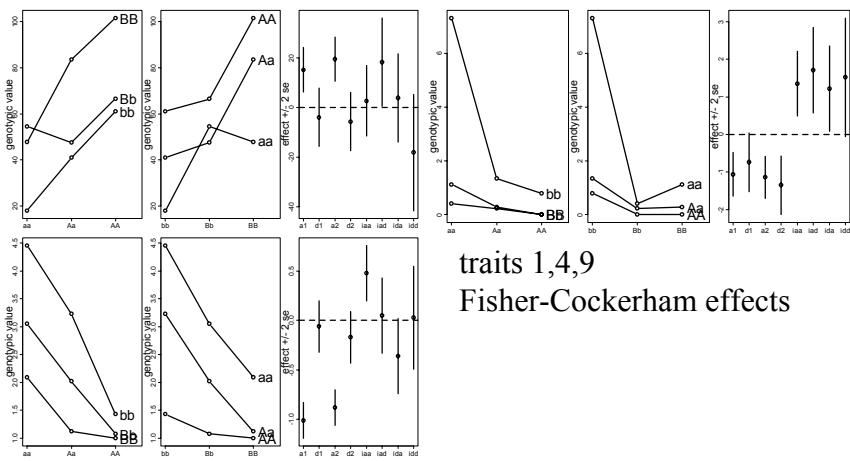
- normal "bell-shaped" environmental variation
- genotypic value G_Q is composite of m QTL
- genetic uncorrelated with environment



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epistasis examples

(Doebley Stec Gustus 1995; Zeng pers. comm.)



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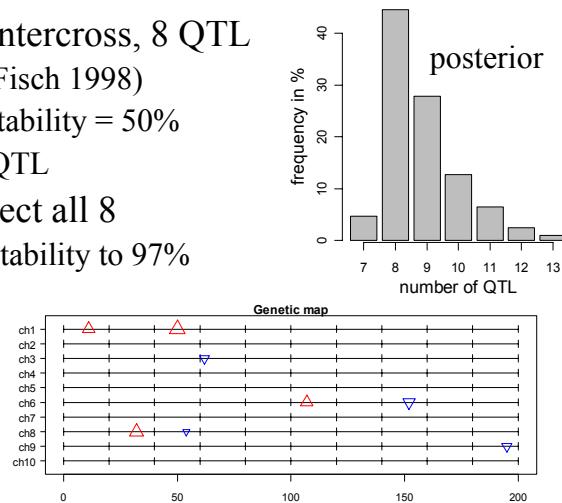
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a complicated simulation

- simulated F2 intercross, 8 QTL
 - (Stephens, Fisch 1998)
 - $n=200$, heritability = 50%
 - detected 3 QTL
- increase to detect all 8
 - $n=500$, heritability to 97%

QTL	chr	loci	effect
1	1	11	-3
2	1	50	-5
3	3	62	+2
4	6	107	-3
5	6	152	+3
6	8	32	-4
7	8	54	+1
8	9	195	+2



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loci pattern across genome

- notice which chromosomes have persistent loci
- best pattern found 42% of the time

Chromosome

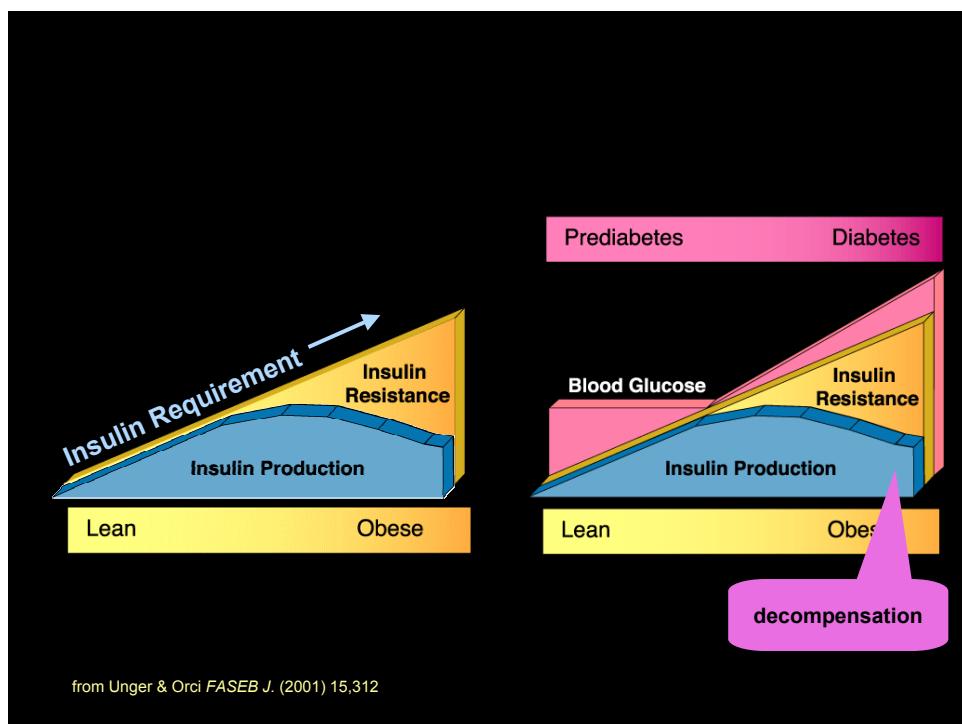
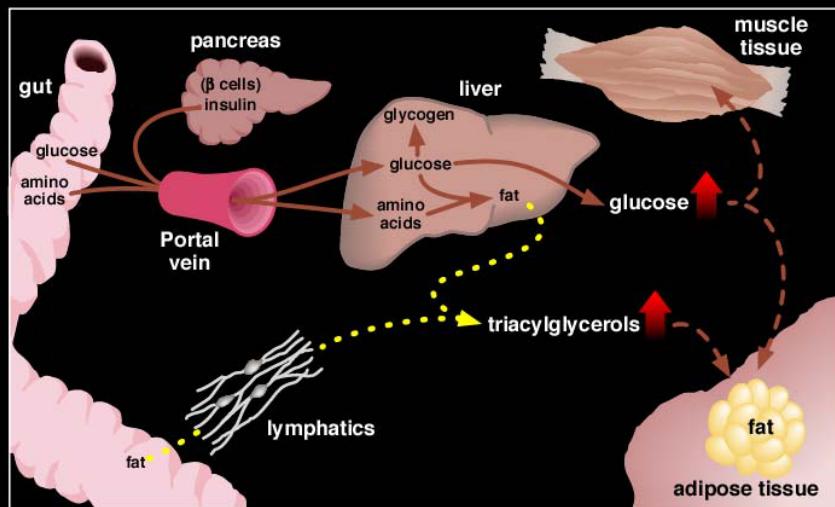
<u>m</u>	1	2	3	4	5	6	7	8	9	10	Count of 8000
8	2	0	1	0	0	2	0	2	1	0	3371
9	3	0	1	0	0	2	0	2	1	0	751
7	2	0	1	0	0	2	0	1	1	0	377
9	2	0	1	0	0	2	0	2	1	0	218
9	2	0	1	0	0	3	0	2	1	0	218
9	2	0	1	0	0	2	0	2	2	0	198

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Type 2 Diabetes Mellitus



from Unger & Orci FASEB J. (2001) 15,312

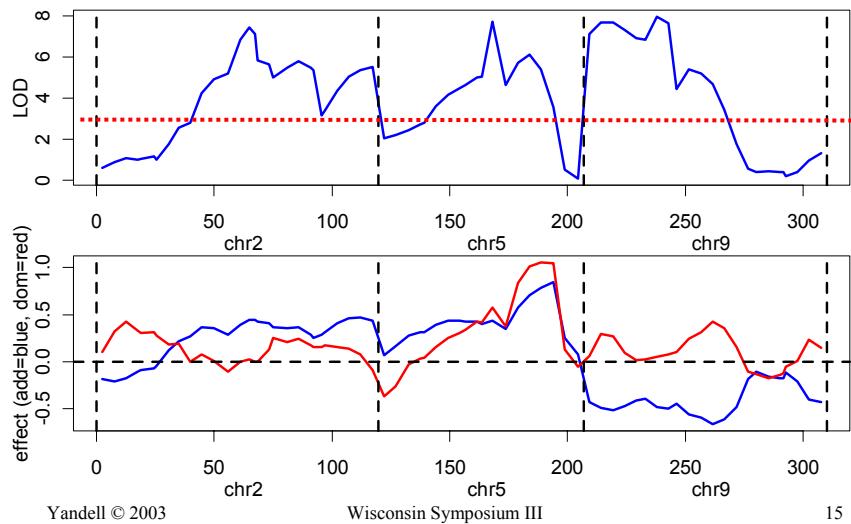
studying diabetes in an F2

- segregating cross of inbred lines
 - B6.ob x BTBR.ob → F1 → F2
 - selected mice with ob/ob alleles at leptin gene (chr 6)
 - measured and mapped body weight, insulin, glucose at various ages (Stoehr et al. 2000 Diabetes)
 - sacrificed at 14 weeks, tissues preserved
- gene expression data
 - Affymetrix microarrays on parental strains, F1
 - key tissues: adipose, liver, muscle, β-cells
 - novel discoveries of differential expression (Nadler et al. 2000 PNAS; Lan et al. 2002 in review; Ntambi et al. 2002 PNAS)
 - RT-PCR on 108 F2 mice liver tissues
 - 15 genes, selected as important in diabetes pathways
 - SCD1, PEPCK, ACO, FAS, GPAT, PPARgamma, PPARalpha, G6Pase, PDI,...

why map gene expression as a quantitative trait?

- *cis-* or *trans*-action?
 - does gene control its own expression?
 - evidence for both modes (Brem et al. 2002 Science)
- mechanics of gene expression mapping
 - measure gene expression in intercross (F2) population
 - map expression as quantitative trait (QTL technology)
 - adjust for multiple testing via false discovery rate
- research groups working on expression QTLs
 - review by Cheung and Spielman (2002 *Nat Gen Suppl*)
 - Kruglyak (Brem et al. 2002 *Science*)
 - Doerge et al. (Purdue); Jansen et al. (Wageningen)
 - Williams et al. (U KY); Lusis et al. (UCLA)
 - Dumas et al. (2000 *J Hypertension*)

Multiple Interval Mapping SCD1: multiple QTL plus epistasis!

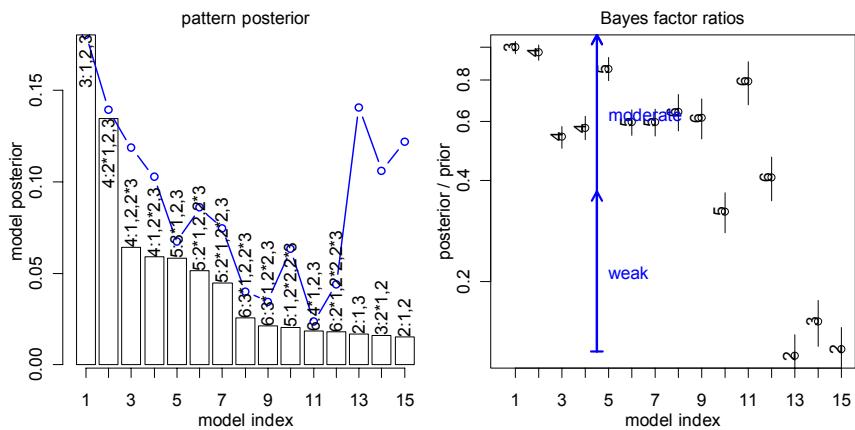


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Bayesian model assessment: chromosome QTL pattern for SCD1



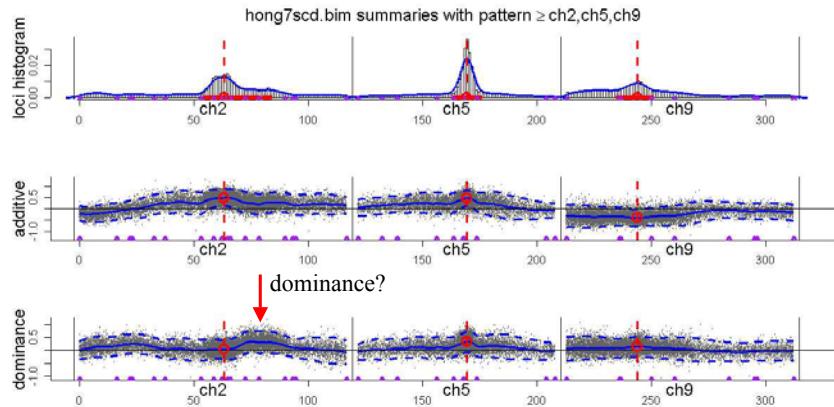
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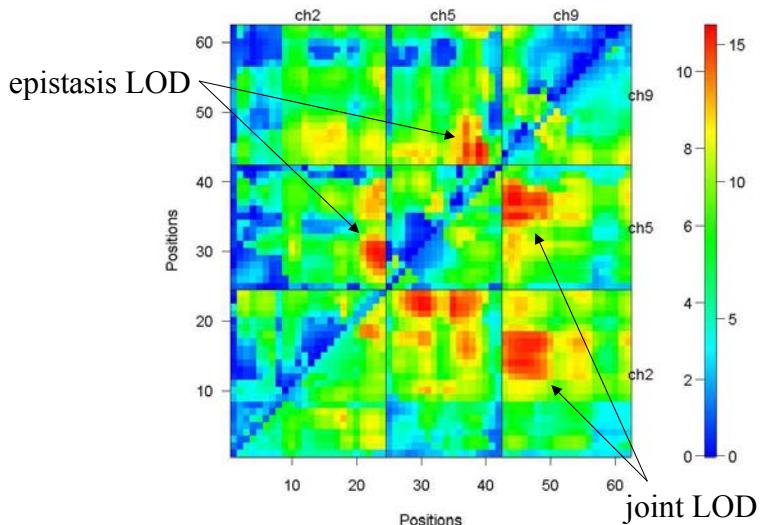
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trans-acting QTL for SCD1

(no epistasis yet: see Yi, Xu, Allison 2003)



2-D scan: assumes only 2 QTL!



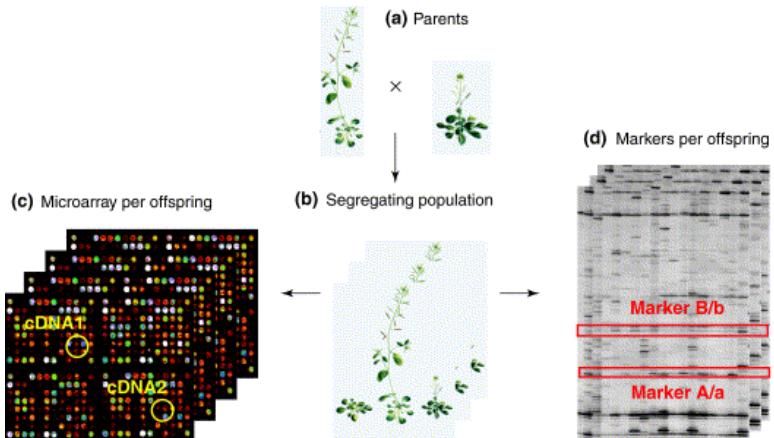
why study multiple traits together?

- environmental correlation
 - non-genetic, controllable by design
 - historical correlation (learned behavior)
 - physiological correlation (same body)
- genetic correlation
 - pleiotropy
 - one gene, many functions
 - common biochemical pathway, splicing variants
 - close linkage
 - two tightly linked genes
 - genotypes Q are collinear

high throughput dilemma

- want to focus on gene expression network
 - ideally capture pathway in a few dimensions
 - allow for complicated genetic architecture
- may have multiple controlling loci
 - could affect many genes in coordinated fashion
 - could show evidence of epistasis
 - quick assessment via interval mapping may be misleading
- try mapping principle components as super-trait
 - capture key multivariate features of multiple traits
 - elicit biochemical pathways (Henderson et al. Hoeschele 2001; Ong Page 2002)

idea of mapping microarrays (Jansen Nap 2001)



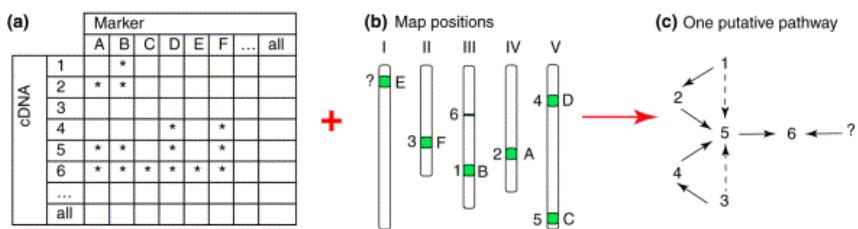
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goal: unravel biochemical pathways (Jansen Nap 2001)

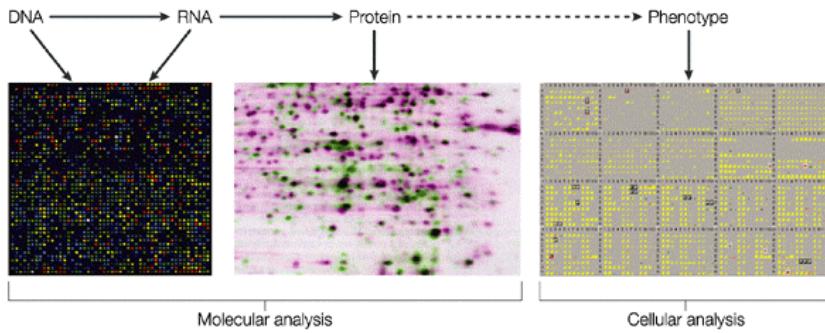


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central dogma via microarrays (Bochner 2003)



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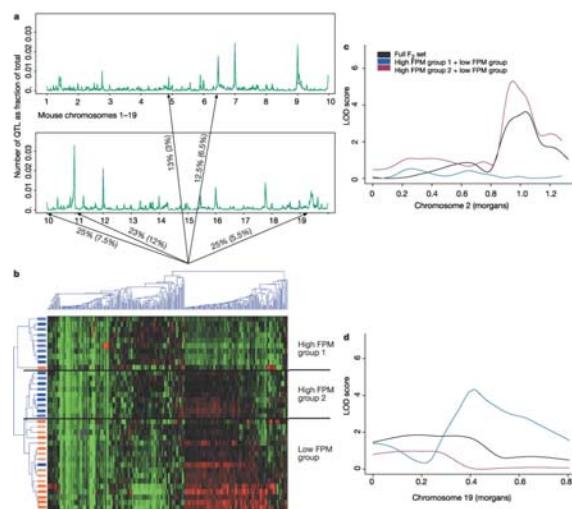
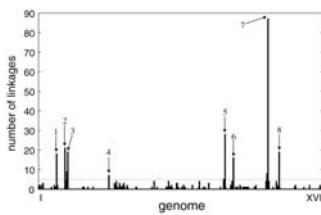
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coordinated expression in mouse genome (Schadt et al. 2003)

expression pleiotropy in yeast genome (Brem et al. 2002)

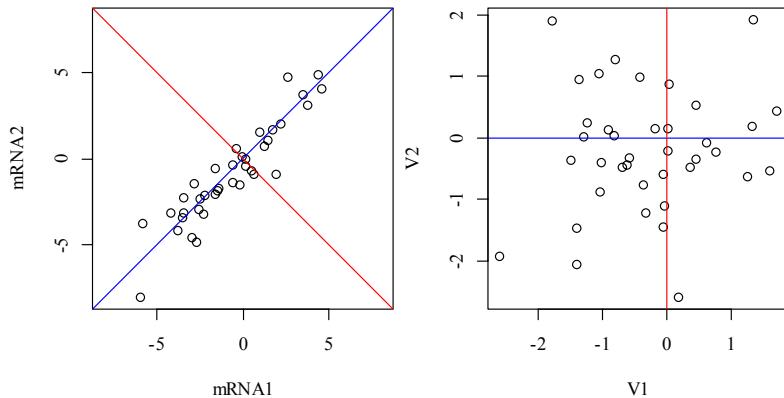


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PC simply rotates & rescales
to find major axes of variation

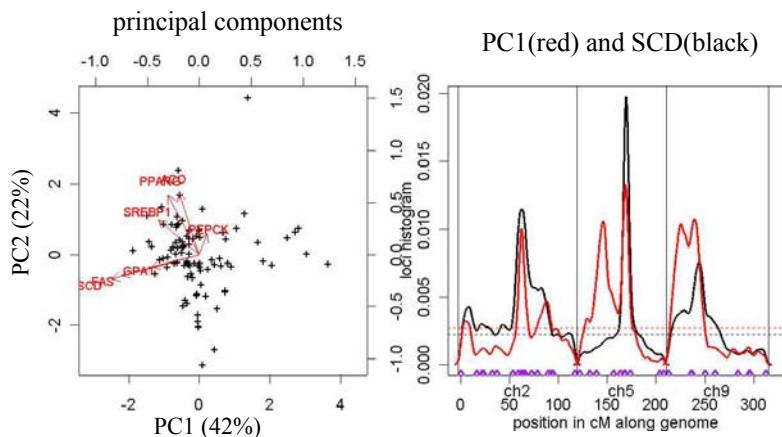


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multivariate screen
for gene expressing mapping

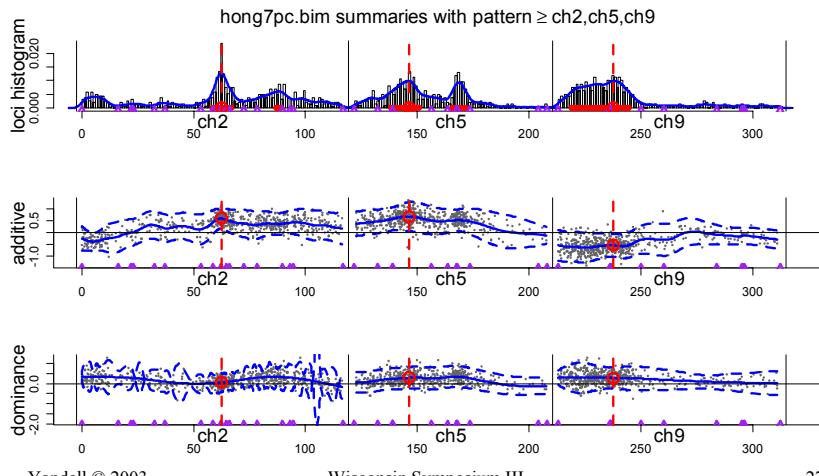


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mapping first diabetes PC as a trait



pFDR for PC1 analysis

